



Read Highlighted Changes: Revised February 2015.

Package insert instructions must be carefully followed. Reliability of assay results cannot be guaranteed if there are any deviations from the instructions in this package insert.

NAME

ARCHITECT Anti-TPO

INTENDED USE

ARCHITECT Anti-TPO is a Chemiluminescent Microparticle Immunoassay (CMIA) for the quantitative determination of the IgG class of thyroid peroxidase autoantibodies (anti-TPO) in human serum and plasma on the ARCHITECT iSystem. The ARCHITECT Anti-TPO assay is intended for use as an aid in the diagnosis of thyroid disease.

SUMMARY AND EXPLANATION OF THE TEST

It was first demonstrated by Trotter et al. in 1957¹ and subsequently by Roitt and Doniach in 1958² that many patients with Hashimoto's thyroiditis had detectable autoantibodies in their blood directed at a thyroid antigen distinct from thyroglobulin. This antigen was termed thyroid microsomal and it has since been demonstrated that most if not all anti-thyroid microsomal autoantibodies recognize thyroid peroxidase (TPO).³

TPO is a membrane-bound glycoprotein enzyme with an approximate mass of 107kD. The *in vivo* function is the iodination of tyrosine in the synthesis of T₃ and T₄.⁴ Autoimmune reactivity to TPO is believed to be polyclonal and heterogeneous in nature with a minimum of six antigenic determinants being recognized, comprising both conformational and linear epitopes.^{5, 6} In addition, the proportion of each immunoglobulin class (G or M) or subclass (G1 – G4) as well as their affinity varies widely from patient to patient.^{7, 8} Unlike autoantibodies to thyroglobulin (anti-Tg), autoantibodies to TPO fix complement,⁹ are potentially deleterious and may have a pathogenic role in (destructive) autoimmune thyroid disease.^{10, 11} Anti-TPO antibodies are found often in conjunction with anti-Tg in the majority of cases of Hashimoto's thyroiditis, Primary Myxedema, and Graves' disease. The relationship of autoimmune thyroid disease to pregnancy has been the subject of considerable interest with the recognition of the postpartum thyroid disease syndromes.¹² Anti-TPO antibodies are demonstrable in most cases of postpartum thyroiditis and it has been found that the presence of autoantibody in early pregnancy was associated with a high risk of asymptomatic postpartum hypothyroidism.¹³⁻¹⁷

It is common to find anti-TPO antibodies in the absence of autoantibodies to thyroglobulin, particularly in patients with small goitres and up to 64% of cases of autoimmune hypothyroidism have been reported to be associated with anti-TPO antibodies alone.¹⁸ In addition, anti-TPO antibodies are frequently found in patients with other autoimmune diseases such as Rheumatoid Arthritis, Addison's Disease and Type I Diabetes.¹⁹⁻²¹ They are also detectable at low levels in up to 20% of asymptomatic individuals,²² particularly the elderly²³ and more often in women than in men, although the clinical significance of these autoantibodies is unclear.

BIOLOGICAL PRINCIPLES OF THE PROCEDURE

The ARCHITECT Anti-TPO assay is a two-step immunoassay for the quantitative determination of anti-TPO in human serum and plasma using CMIA technology with flexible assay protocols, referred to as Chemiflex.

1. Sample, assay diluent, and TPO coated paramagnetic microparticles are combined and incubated. The anti-TPO present in the sample binds to the TPO coated microparticles.
2. After washing, anti-human IgG acridinium-labeled conjugate is added to create a reaction mixture.
3. Following another incubation and wash, Pre-Trigger and Trigger Solutions are added to the reaction mixture.
4. The resulting chemiluminescent reaction is measured as relative light units (RLUs). There is a direct relationship between the amount of anti-TPO in the sample and the RLUs detected by the ARCHITECT iSystem optics.


For additional information on system and assay technology, refer to the ARCHITECT System Operations Manual, Section 3.

REAGENTS

Kit Contents

ARCHITECT Anti-TPO 2K47

NOTE: Some kit sizes are not available in all countries or for use on all ARCHITECT iSystems. Please contact your local distributor.

REF	2K47-25	2K47-20
	100	400
MICROPARTICLES	1 x 6.6 mL	4 x 6.6 mL
CONJUGATE	1 x 5.9 mL	4 x 5.9 mL
ASSAY DILUENT	1 x 10.0 mL	4 x 10.0 mL
MICROPARTICLES	Thyroid peroxidase (recombinant) coated microparticles in MES buffer with protein (bovine) stabilizer. Minimum concentration: 0.10 % solids. Preservative: antimicrobial agents.	
CONJUGATE	Anti-human IgG (mouse, monoclonal) acridinium-labeled conjugate in MES buffer with protein (bovine) stabilizer. Minimum concentration: 80.0 ng/mL. Preservative: antimicrobial agents.	
ASSAY DILUENT	Assay Diluent in MES buffer. Preservative: antimicrobial agents.	

Other Reagents

PRE-TRIGGER SOLUTION ARCHITECT Pre-Trigger Solution containing 1.32% (w/v) hydrogen peroxide.

TRIGGER SOLUTION ARCHITECT Trigger Solution containing 0.35 N sodium hydroxide.


WASH BUFFER ARCHITECT Wash Buffer containing phosphate buffered saline solution. Preservatives: antimicrobial agents.


Warnings and Precautions

- **IVD**
- For *In Vitro* Diagnostic Use

Safety Precautions

CAUTION: This product requires the handling of human specimens. It is recommended that all human-sourced materials be considered potentially infectious and handled in accordance with the OSHA Standard on Bloodborne Pathogens. Biosafety Level 2 or other appropriate biosafety practices should be used for materials that contain or are suspected of containing infectious agents.²⁴⁻²⁷

The following warnings and precautions apply to: MICROPARTICLES	
	
WARNING	Contains potassium ferricyanide.
H361	Suspected of damaging fertility or the unborn child.
Prevention	
P201	Obtain special instructions before use.
P280	Wear protective gloves / protective clothing / eye protection.
Response	
P308+P313	IF exposed or concerned: Get medical advice / attention.
Disposal	
P501	Dispose of contents / container in accordance with local regulations.

The following warnings and precautions apply to: ASSAY DILUENT	
	
WARNING	
H319	Causes serious eye irritation.
Prevention	
P264	Wash hands thoroughly after handling.
P280	Wear protective gloves / protective clothing / eye protection.
Response	
P305+P351+P338	IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.
P337+P313	If eye irritation persists: Get medical advice / attention.

Safety Data Sheets are available at www.abbottiagnostics.com or contact your local representative.

For a detailed discussion of safety precautions during system operation, refer to the ARCHITECT System Operations Manual, Section 8.

Reagent Handling

- Do not use reagent kits beyond the expiration date.
- Do not pool reagents within a kit or between kits.**
- Before loading the reagent kit on the system for the first time, the microparticle bottle requires mixing to resuspend microparticles that may have settled during shipment. For microparticle mixing instructions, refer to the **PROCEDURE, Assay Procedure** section of this package insert.
- Septums MUST be used to prevent reagent evaporation and contamination and to ensure reagent integrity. Reliability of assay results cannot be guaranteed if septums are not used according to the instructions in this package insert.**
 - To avoid contamination, wear clean gloves when placing a septum on an uncapped reagent bottle.**
 - Once a septum has been placed on an open reagent bottle, **do not invert the bottle** as this will result in reagent leakage and may compromise assay results.
 - Over time, residual liquids may dry on the septum surface. These are typically dried salts and have no effect on assay efficacy.

For a detailed discussion of handling precautions during system operation, refer to the ARCHITECT System Operations Manual, Section 7.

Reagent Storage

When stored and handled as directed, reagents are stable until the expiration date.

	Storage Temperature	Maximum Storage Time	Additional Storage Instructions
Unopened/ Opened*	2-8°C	Until expiration date	May be used immediately after removal from 2-8°C storage. Store in upright position.
On board	System temperature	30 days	Discard after 30 days. For information on tracking onboard time, refer to the ARCHITECT System Operations Manual, Section 5.

* Reagents may be stored on or off the ARCHITECT iSystem. If reagents are removed from the system, store them at 2-8°C (with septums and replacement caps) in an upright position. For reagents stored off the system, it is recommended that they be stored in their original trays and boxes to ensure they remain upright. **If the microparticle bottle does not remain upright (with a septum installed) while in refrigerated storage off the system, the reagent kit must be discarded.** For information on unloading reagents, refer to the ARCHITECT System Operations Manual, Section 5.

After reagents are removed from the system, one must initiate a scan to update the onboard stability timer.

Indications of Reagent Deterioration

When a control value is out of the specified range, it may indicate deterioration of the reagents or errors in technique. Associated test results are invalid, and samples must be retested. Assay recalibration may be necessary. For troubleshooting information, refer to the ARCHITECT System Operations Manual, Section 10.

INSTRUMENT PROCEDURE

The ARCHITECT Anti-TPO assay file must be installed on the ARCHITECT iSystem prior to performing the assay. For detailed information on assay file installation and viewing and editing assay parameters, refer to the ARCHITECT System Operations Manual, Section 2.

For information on printing assay parameters, refer to the ARCHITECT System Operations Manual, Section 5.

For a detailed description of system procedures, refer to the ARCHITECT System Operations Manual.

The default result unit for the ARCHITECT Anti-TPO assay is IU/mL.

SPECIMEN COLLECTION AND PREPARATION FOR ANALYSIS

Specimen Types

Verified specimen types to be used with this assay:

Specimen Types	Collection Tubes	
	Glass	Plastic
Serum	No additive (uncoated)	Serum separator tubes
Plasma	Lithium heparin	Lithium heparin
	Plasma separator tubes with lithium heparin	Plasma separator tubes with lithium heparin
	EDTA	Sodium heparin EDTA

- Other specimen collection tube types have not been tested with this assay.
- When serial specimens are being evaluated, the same type of specimen should be used throughout the study.
- Performance has not been established for the use of cadaveric specimens or the use of body fluids other than human serum or plasma.

- The instrument does not provide the capability to verify specimen type. It is the responsibility of the operator to verify that the correct specimen types are used in the assay.

Specimen Conditions

- Do not use specimens with the following conditions:
 - heat-inactivated
 - obvious microbial contamination
- For accurate results, serum and plasma specimens should be free of fibrin, red blood cells, and other particulate matter. Serum specimens from patients receiving anticoagulant or thrombolytic therapy may contain fibrin due to incomplete clot formation.
- Ensure that complete clot formation in serum specimens has taken place prior to centrifugation. Some specimens, especially those from patients receiving anticoagulant or thrombolytic therapy, may exhibit increased clotting time. If the specimen is centrifuged before a complete clot forms, the presence of fibrin may cause erroneous results.
- To prevent cross contamination, use of disposable pipettes or pipette tips is recommended.

Preparation for Analysis

- Follow the tube manufacturer’s processing instructions for specimen collection tubes.
- Patient specimens with a cloudy or turbid appearance must be centrifuged prior to testing. Following centrifugation, avoid the lipid layer (if present) when pipetting the specimen into a sample cup or secondary tube.
- Specimens must be mixed THOROUGHLY after thawing by LOW speed vortexing or by gently inverting, and centrifuged prior to use to remove red blood cells or particulate matter to ensure consistency in the results. Multiple freeze-thaw cycles of specimens should be avoided.
- All samples (patient specimens, controls, and calibrators) should be tested within 3 hours of being placed on board the ARCHITECT System. Refer to the ARCHITECT System Operations Manual, Section 5, for a more detailed discussion of onboard sample storage constraints.
- Inspect all specimens for bubbles. Remove bubbles with an applicator stick before analysis. Use a new applicator stick for each specimen to prevent cross contamination.

Specimen Storage

Specimen Type	Storage Temperature	Maximum Storage Time
Serum/Plasma	Room temperature	≤ 8 hours
	2-8°C	≤ 72 hours
	-10°C or colder	≤ 30 days

If testing will be delayed for more than 8 hours, remove serum or plasma from the serum or plasma separator, red blood cells or clot. Specimens removed from the separator gel, cells or clot may be stored up to 72 hours at 2-8°C.

Specimens can be stored up to 30 days frozen at -10°C or colder.

Specimen Shipping

- Package and label specimens in compliance with applicable state, federal, and international regulations covering the transport of clinical specimens and infectious substances.
- It is recommended that specimens be removed from the clot, red blood cells, or separator gel.
- Specimens may be shipped ambient for up to 8 hours after collection or on wet or dry ice for up to 72 hours after collection.
- Do not exceed the storage time limitations listed above.

PROCEDURE

Materials Provided

2K47 ARCHITECT Anti-TPO Reagent Kit

Materials Required but not Provided

- ARCHITECT Anti-TPO Assay file obtained from the ARCHITECT iSystem e-Assay CD-ROM found on www.abbottdiagnostics.com.
- 2K47-01 ARCHITECT Anti-TPO Calibrators
- 2K47-10 ARCHITECT Anti-TPO Controls
- ARCHITECT Pre-Trigger Solution
- ARCHITECT Trigger Solution
- ARCHITECT Wash Buffer
- ARCHITECT Reaction Vessels
- ARCHITECT Sample Cups
- ARCHITECT Septum
- ARCHITECT Replacement Caps
- Pipettes or pipette tips (optional) to deliver the volumes specified on the patient or control order screen.

For information on materials required for maintenance procedures, refer to the ARCHITECT System Operations Manual, Section 9.

Assay Procedure

- Before loading the reagent kit on the system for the first time, the microparticle bottle requires mixing to resuspend microparticles that may have settled during shipment. After the first time the microparticles have been loaded, no further mixing is required.
 - **Invert the microparticle bottle 30 times.**
 - Visually inspect the bottle to ensure microparticles are resuspended. If microparticles are still adhered to the bottle, continue to invert the bottle until the microparticles have been completely resuspended.
 - **If the microparticles do not resuspend, DO NOT USE. Contact your local Abbott representative.**
 - Once the microparticles have been resuspended, place a septum on the bottle. For instructions about placing septums on bottles, refer to the **Reagent Handling** section of this package insert.
- Load the reagent kit on the ARCHITECT iSystem.
 - Verify that all necessary reagents are present.
 - Ensure that septums are present on all reagent bottles.
- Order calibration, if necessary.
 - For information on ordering calibrations, refer to the ARCHITECT System Operations Manual, Section 6.
- Order tests.
 - For information on ordering patient specimens and controls and for general operating procedures, refer to the ARCHITECT System Operations Manual, Section 5.
- Minimum sample cup volume is calculated by the system and printed on the Orderlist report. Verify adequate sample cup volume is present prior to running the test.
 - Maximum number of replicates sampled from the same sample cup: 10
 - Priority:
 - Sample volume for first test: 75 µL
 - Sample volume for each additional test from same sample cup: 25 µL
 - ≤ 3 hours on board:
 - Sample volume for first test: 150 µL
 - Sample volume for each additional test from same sample cup: 25 µL
 - To minimize the effects of evaporation, all samples (patient specimens, calibrators and controls) must be tested within 3 hours of being placed on board the ARCHITECT iSystem.
 - If using primary or aliquot tubes, use the sample gauge to ensure sufficient patient specimen is present.
- Prepare ARCHITECT Anti-TPO Calibrators and Controls.

- ARCHITECT Anti-TPO Calibrators and Controls should be prepared according to their respective package inserts.
- Hold bottles **vertically** and dispense recommended volumes into each respective sample cup.
- Recommended volumes:
for each calibrator: 5 drops
for each control: 5 drops
- Load samples.
 - For information on loading samples, refer to the ARCHITECT System Operations Manual, Section 5.
- Press RUN. The system performs the following functions:
 - Moves the sample to the aspiration point.
 - Loads a reaction vessel (RV) into the process path.
 - Aspirates and transfers sample into the RV.
 - Advances the RV one position and transfers assay diluent and microparticles into the RV.
 - Mixes, incubates, and washes the reaction mixture.
 - Adds conjugate to the RV.
 - Mixes, incubates, and washes the reaction mixture.
 - Adds pre-trigger and trigger solutions.
 - Measures chemiluminescent emission to determine the quantity of anti-TPO in the sample.
 - Aspirates contents of RV to liquid waste and unloads RV to solid waste.
 - Calculates the result.
- For additional information on principles of operation, refer to the ARCHITECT System Operations Manual, Section 3.
- For optimal performance, it is important to perform routine maintenance as described in the ARCHITECT System Operations Manual, Section 9. Perform maintenance more frequently when required by laboratory procedures.

Specimen Dilution Procedures

Specimens with an anti-TPO value exceeding 1000.00 IU/mL are flagged with the code "> 1000.00" and may be diluted using the Automated Dilution Protocol or the Manual Dilution Procedure.

Automated Dilution Protocol

The system performs a 1:2 dilution of the specimen and automatically calculates the concentration of the specimen before dilution and reports the result.

Specimens with an anti-TPO value exceeding 2000.00 IU/mL are flagged with the code "> 2000.00" and may be diluted using the Manual Dilution Procedure.

Manual Dilution Procedure

Suggested dilution: 1:20

1. Prior to diluting the specimen, dispense approximately 10 drops of ARCHITECT Anti-TPO Calibrator A into a clean test tube for use in the next step.
2. Transfer 190 µL of ARCHITECT Anti-TPO Calibrator A from the test tube prepared in the prior step into another clean test tube and add 10 µL of the patient specimen.
3. The operator must enter the dilution factor in the Patient or Control order screen. The system will use this dilution factor to automatically calculate the concentration of the sample before dilution and report the result. The result should be > 5.61 IU/mL before the dilution factor is applied.

For detailed information on ordering dilutions, refer to the ARCHITECT System Operations Manual, Section 5.

Calibration

- Test Calibrators A-F in duplicate. The calibrators should be priority loaded.
A single sample of each control level must be tested to evaluate the assay calibration. Ensure that assay control values are within the ranges specified in the respective control package insert.

- Calibration Range: 0.00 - 1000.00 IU/mL.
- Once an ARCHITECT Anti-TPO calibration is accepted and stored, all subsequent samples may be tested without further calibration unless:
 - A reagent kit with a new lot number is used or
 - Controls are out of range.
- For detailed information on how to perform an assay calibration, refer to the ARCHITECT System Operations Manual, Section 6.

Quality Control Procedures

The recommended control requirement for the ARCHITECT Anti-TPO assay is that a single sample of each control level be tested once every 24 hours each day of use. If the quality control procedures in your laboratory require more frequent use of controls to verify test results, follow your laboratory-specific procedures.

The ARCHITECT Anti-TPO Control values must be within the acceptable ranges specified in the control package insert. If a control is out of its specified range, the associated test results are invalid and must be retested. Recalibration may be indicated.

Verification of Assay Claims

For protocols to verify package insert claims, refer to the ARCHITECT System Operations Manual, Appendix B.

The ARCHITECT Anti-TPO assay belongs to method group 1.

RESULTS

Calculation

The ARCHITECT Anti-TPO assay utilizes a 4 Parameter Logistic Curve fit data reduction method (4PLC, Y-weighted) to generate a calibration curve.

Flags

Some results may contain information in the Flags field. For a description of the flags that may appear in this field, refer to the ARCHITECT System Operations Manual, Section 5.

LIMITATIONS OF THE PROCEDURE

- Antibody measurement represents one parameter in a multi-criteria diagnostic process. When making a diagnosis of thyroid disease, a combination of test methods should be used in conjunction with clinical symptoms.
- About 20% of asymptomatic specimens may present with anti-TPO autoantibodies reflecting the prevalence in apparently healthy populations. The prevalence of anti-TPO may also depend on age, gender, and geographic region of the selected population.
- Some specimens may not dilute linearly because of the heterogeneity of the autoantibodies with respect to physicochemical properties.
- Specimens from patients who have received preparations of mouse monoclonal antibodies for diagnosis or therapy may contain human anti-mouse antibodies (HAMA). Such specimens may show either falsely elevated or depressed values when tested with assay kits that employ mouse monoclonal antibodies. Assay results that are not consistent with other clinical observations may require additional information for diagnosis.^{28, 29}
- Heterophilic antibodies in human serum can react with reagent immunoglobulins, interfering with *in vitro* immunoassays. The presence of heterophilic antibodies in a patient specimen may cause anomalous values to be observed. Additional information may be required for diagnosis.³⁰

EXPECTED VALUES

In a study, human serum specimens were collected from a population of 236 apparently healthy individuals. All specimens delivered TSH values within the normal reference range. Of this study population, 9 specimens delivered positive results on a commercially available anti-TPO assay device and were excluded from further normal range analysis. The 97.5 percentile concentration of the remaining population was 5.61 IU/mL. In this study population, the normal range

is < 5.61 IU/mL. A total of 97.8% (222/227) of the population gave values within this normal range.* This normal range is suggested as a guideline and each laboratory should establish a normal range appropriate to their patient populations, giving due consideration to age, gender, geographical location and their clinical practice.

* Representative data; results in individual laboratories may vary from these data.

SPECIFIC PERFORMANCE CHARACTERISTICS

Precision

The ARCHITECT Anti-TPO assay is designed to have an assay precision of $\leq 10\%$ total CV for samples ≥ 5.61 IU/mL.

A study was performed for the ARCHITECT Anti-TPO assay with guidance from the National Committee for Clinical Laboratory Standards (NCCLS) Protocol EP5-A.³¹ ARCHITECT Anti-TPO Positive Control and three human panels were assayed using three lots of reagents in replicates of two at two separate times per day for 20 days on three instruments. Each reagent lot used a single calibration curve throughout the study. Data from this study are summarized in the following table.*

Sample	Instrument	Reagent Lot	n	Mean	Within Run		Total	
				Conc. Value (IU/mL)	SD	%CV	SD	%CV
Positive Control	1	A	80	74.85	2.11	2.8	2.16	2.9
		B	80	74.20	1.89	2.6	2.04	2.7
		C	80	74.63	2.01	2.7	2.17	2.9
	2	A	80	77.42	2.11	2.7	3.25	4.2
		B	80	75.32	1.92	2.5	2.54	3.4
		C	80	74.59	1.73	2.3	2.57	3.4
	3	A	80	75.41	2.13	2.8	2.52	3.3
		B	80	75.48	1.90	2.5	2.13	2.8
		C	80	76.66	2.42	3.2	2.87	3.7
Panel 1	1	A	80	1.57	0.08	4.8	0.10	6.5
		B	80	1.46	0.06	3.8	0.09	5.8
		C	80	1.64	0.09	5.6	0.10	6.1
	2	A	80	1.60	0.09	5.3	0.12	7.6
		B	80	1.53	0.06	3.9	0.11	7.2
		C	80	1.52	0.10	6.7	0.12	7.7
	3	A	80	1.47	0.08	5.3	0.11	7.8
		B	80	1.47	0.07	4.7	0.13	8.5
		C	80	1.52	0.14	9.5	0.15	9.8
Panel 2	1	A	80	20.98	0.65	3.1	0.76	3.6
		B	80	21.14	0.61	2.9	0.66	3.1
		C	80	21.51	0.71	3.3	0.75	3.5
	2	A	80	21.27	0.61	2.9	0.98	4.6
		B	80	21.62	0.66	3.0	0.90	4.2
		C	80	20.82	0.67	3.2	0.85	4.1
	3	A	80	21.00	0.73	3.5	0.86	4.1
		B	80	21.77	0.60	2.7	0.84	3.8
		C	80	21.24	0.70	3.3	0.89	4.2
Panel 3	1	A	80	214.78	5.14	2.4	6.48	3.0
		B	80	221.79	4.73	2.1	5.82	2.6
		C	80	216.71	5.36	2.5	6.36	2.9
	2	A	80	219.32	4.41	2.0	8.61	3.9
		B	80	224.54	4.04	1.8	13.37	6.0
		C	80	218.73	5.76	2.6	13.18	6.0
	3	A	80	212.91	6.11	2.9	6.84	3.2
		B	80	225.46	5.15	2.3	5.67	2.5
		C	80	228.17	5.80	2.5	7.09	3.1

* Representative data; results in individual laboratories may vary from these data.

Functional Sensitivity

In a study, human panels ranging in concentration from 0.16-1.20 IU/mL were tested in replicates of 2 over 10 days on one instrument using two reagent lots and three calibrations for a total of 40 replicates per panel. The total %CVs (combining variance components for replicate, run, day and reagent lot) were calculated and plotted against the mean concentration. A reciprocal curve was fitted through the data and the functional sensitivity value was calculated as the concentration corresponding to the 20% CV level of the fitted curve. The lowest ARCHITECT Anti-TPO assay value exhibiting a 20% CV is 0.50 IU/mL.*

* Representative data; results in individual laboratories may vary from these data.

Analytical Sensitivity

The ARCHITECT Anti-TPO assay is designed to have an analytical sensitivity of ≤ 1.0 IU/mL. The analytical sensitivity of the ARCHITECT Anti-TPO assay, defined as the concentration at two standard deviations above the ARCHITECT Anti-TPO Calibrator A (0.0 IU/mL) was calculated to be 0.16 IU/mL* at the 95% level of confidence (based upon one study with n=48 runs, 10 replicates of Calibrator A and 4 replicates of Calibrator B per run).

* Representative data; results in individual laboratories may vary from these data.

Linearity

The ARCHITECT Anti-TPO assay is linear between 3.0 and 1000.0 IU/mL based on a study performed with guidance from NCCLS protocol EP6-A.³²

Autodilution Verification

The ARCHITECT Anti-TPO automated dilution protocol is designed to recover within 15% of manually diluted specimens. In a study, the automated dilution protocol (1:2) was compared to a manual 1:2 dilution procedure using 9 human specimens with anti-TPO levels that were greater than Calibrator E (250 IU/mL). The manual dilution was performed with ARCHITECT Anti-TPO Calibrator A. The observed percent recovery results are summarized in the following table.*

Sample ID	Automated Dilution (IU/mL)	Manual Dilution (IU/mL)	% Recovery**
1	861.25	859.27	100.2
2	684.49	703.64	97.3
3	844.36	847.62	99.6
4	724.55	757.09	95.7
5	709.46	688.49	103.1
6	1105.65	1106.18	100.0
7	948.43	931.80	101.8
8	840.77	851.72	98.7
9	966.48	998.45	96.8

* Representative data; results in individual laboratories may vary from these data.

$$** \% \text{ Recovery} = \frac{\text{Automated Dilution (IU/mL)}}{\text{Manual Dilution (IU/mL)}} \times 100$$

Interference

Interference from elevated levels of bilirubin, hemoglobin, triglycerides, and total protein in the ARCHITECT Anti-TPO assay is designed to be $\leq 15\%$ at the levels indicated.

A study based on guidance from the NCCLS Protocol EP7-A³³ was performed for the ARCHITECT Anti-TPO assay. Specimens with anti-TPO levels between 45.07 and 361.64 IU/mL were supplemented with the following potentially interfering compounds. The average amount of interference observed during the study ranged from -3.6% to +3.7%.*

Potentially Interfering Substance	Potentially Interfering Substance Concentration
Bilirubin	20 mg/dL
Hemoglobin	1000 mg/dL
Total Protein (Low)	4 g/dL
Total Protein (High)	10 g/dL
Triglycerides	1000 mg/dL

* Representative data; results in individual laboratories may vary from these data.

Evaluation of Autoimmune Disease Specimens and High Titer IgG Samples

Potential interference from autoimmune disease specimens and high titer IgG samples in the ARCHITECT Anti-TPO assay is designed to be $\leq 15\%$. In a study, the ARCHITECT Anti-TPO assay was evaluated by testing specimens with known autoimmune diseases and elevated IgG. Specimens were evaluated with anti-TPO levels spiked between 131.44 and 568.78 IU/mL. Mean absolute % interference is summarized in the following table.*

Clinical Condition	Mean Absolute % Interference
Anti-Nuclear Antibody (ANA)	1.6
Rheumatoid Arthritis (RA)	1.6
Systemic Lupus Erythematosus (SLE)	1.1
Insulin Dependent Diabetes Mellitus (IDDM)	1.0
Crohn's Disease	2.4
Multiple Sclerosis	1.7
Ulcerative Colitis	1.5
Hyperglobulinemia (high IgG)	0.9

* Representative data; results in individual laboratories may vary from these data.

Evaluation of Other Potential Interferents

Potential interference from HAMA and rheumatoid factor (RF) in the ARCHITECT Anti-TPO assay is designed to be $\leq 15\%$. In a study, the ARCHITECT Anti-TPO assay was evaluated by testing specimens with HAMA and RF to further assess the clinical specificity. Specimens positive for HAMA and specimens positive for RF were evaluated for % interference with anti-TPO levels spiked between 163.0 and 184.3 IU/mL. Mean absolute % interference is summarized in the following table.*

Other Potential Interferents	Number of Specimens	Mean Absolute % Interference
HAMA Positive	10	2.1
RF Positive	10	1.6

* Representative data; results in individual laboratories may vary from these data.

Clinical Sensitivity

In two studies, clinical sensitivity was evaluated by testing 139 clinically defined Hashimoto's thyroiditis specimens and 125 Graves' disease specimens. The clinical diagnosis was based on the criteria of the respective laboratory. The presence of autoantibodies against thyroglobulin and/or TPO was not necessarily a diagnostic criterion of these Graves' disease and Hashimoto's thyroiditis specimens. Data from these studies are summarized in the following table.*

	Hashimoto's Thyroiditis		Graves' Disease	
	n	% Positive	n	% Positive
Study 1	89	64.0	75	92.0
Study 2	50	74.0	50	100.0

* Representative data; results in individual laboratories may vary from these data.

Concordance

The performance of the ARCHITECT Anti-TPO was compared to a commercially available immunoassay for the determination of anti-TPO. A total of 500 specimens were evaluated in a study, encompassing a population of apparently healthy individuals and patients with autoimmune thyroid disease (Graves' disease and Hashimoto's thyroiditis). Specimens were tested in replicates of one using the ARCHITECT Anti-TPO assay with three reagent lots on three instruments and compared with a commercially available immunoassay (Comparison Assay). Data from this study are summarized in the following table.*

ARCHITECT Anti-TPO	Comparison Assay	
	Negative	Positive
Negative	242	32
Positive	5	221
Concordance = 92.6 %		


* Representative data; results in individual laboratories may vary from these data.


BIBLIOGRAPHY

- Trotter WR, Belyavin G, Waddams A. Precipitating and complement-fixing antibodies in Hashimoto's disease. *Proc Royal Soc Med.* 1957;50:961-962.
- Roitt IM, Doniach D. Human auto-immune thyroiditis: serological studies. *Lancet.* 1958;7055:1027-1033.
- Ruf J, Czarnocka B, De Micco C, et al. Thyroid peroxidase is the organ-specific 'microsomal' autoantigen involved in thyroid autoimmunity. *Acta Endocrinol (Copenh).* 1987;S281:49-56.
- Degroot LJ, Niepomniszcze H. Biosynthesis of thyroid hormone: basic and clinical aspects. *Metabolism.* 1977;26(6):665-718.
- Doble ND, Banga JP, Pope R, et al. Autoantibodies to the thyroid microsomal / thyroid peroxidase antigen are polyclonal and directed to several distinct antigenic sites. *Immunology.* 1988;64:23-29.
- Libert F, Ludgate M, Dinsart C, et al. Thyroperoxidase, but not the thyrotropin receptor, contains sequential epitopes recognized by autoantibodies in recombinant peptides expressed in the pUEX vector. *J Clin Endocrinol Metab.* 1991;73(4):857-860.
- Parkes AB, McLachlan SM, Bird P, et al. The distribution of microsomal and thyroglobulin antibody activity among the IgG subclasses. *Clin Exp Immunol.* 1984;57:239-243.
- Weetman AP, Black CM, Cohen SB, et al. Affinity purification of IgG subclasses and the distribution of thyroid auto-antibody reactivity in Hashimoto's thyroiditis. *Scan J Immunol.* 1989;30:73-82.
- Holborow EJ, Brown PC, Roitt IM, et al. Cytoplasmic localization of "complement-fixing" auto-antigen in human thyroid epithelium. *Br J Exp Pathol.* 1959;XL(6):583-588.
- Khoury EL, Hammond L, Bottazzo GF, et al. Presence of the organ-specific 'microsomal' autoantigen on the surface of human thyroid cells in culture: its involvement in complement-mediated cytotoxicity. *Clin Exp Immunol.* 1981;45:316-328.
- Banga JP, Pryce G, Hammond L, et al. Structural features of the autoantigens involved in thyroid autoimmune disease: the thyroid microsomal / microvillar antigen. *Mol Immunol.* 1985;22(6):629-642.
- Davies TF, Weiss I. Autoimmune thyroid disease and pregnancy. *Am J Reprod Immunol.* 1981;1:187-192.
- Amino N, Yabu Y, Miki T, et al. Serum ratio of triiodothyronine to thyroxine, and thyroxine-binding globulin and calcitonin concentrations in Graves' disease and destruction-induced thyrotoxicosis. *J Clin Endocrinol Metab.* 1981;53(1):113-116.
- Jansson R, Bernander S, Karlsson A, et al. Autoimmune thyroid dysfunction in the postpartum period. *J Clin Endocrinol Metab.* 1984;58(4):681-687.
- Othman S, Phillips DIW, Parkes AB, et al. A long-term follow-up of postpartum thyroiditis. *Clin Endocrinol.* 1990;32:559-564.
- Harris B, Othman S, Davies JA, et al. Association between postpartum thyroid dysfunction and thyroid antibodies and depression. *BMJ.* 1992;305:152-156.
- Glinoe D. The systematic screening and management of hypothyroidism and hyperthyroidism during pregnancy. *TEM.* 1998;9(10):403-411.
- Nordyke RA, Gilbert FI Jr, Miyamoto LA, et al. The superiority of antimicrosomal over antithyroglobulin antibodies for detecting Hashimoto's thyroiditis. *Arch Intern Med.* 1993;153:862-865.
- Chang C-C, Huang C-N, Chuang L-M. Autoantibodies to thyroid peroxidase in patients with type I diabetes in Taiwan. *Eur J Endocrinol.* 1998;139:44-48.
- Walker DJ, Griffiths M, Griffiths ID. Occurrence of autoimmune diseases and autoantibodies in multicase rheumatoid arthritis families. *Ann Rheum Dis.* 1986;45:323-326.
- Scherbaum WA. On the clinical importance of thyroid microsomal and thyroglobulin antibody determination. *Acta Endocrinol (Copenh).* 1987;S281:325-329.
- Rosenbaum D, Davies TF. The clinical use of thyroid autoantibodies. *The Endocrinologist.* 1992;2(1):55-62.

23. Mariotti S, Chiovato L, Franceschi C, et al. Thyroid autoimmunity and aging. *Exp Gerontol*. 1998;33(6):535-541.
24. US Department of Labor, Occupational Safety and Health Administration, 29 CFR Part 1910.1030, Bloodborne pathogens.
25. US Department of Health and Human Services. *Biosafety in Microbiological and Biomedical Laboratories*. 5th ed. Washington, DC: US Government Printing Office; December 2009.
26. World Health Organization. *Laboratory Biosafety Manual*. 3rd ed. Geneva: World Health Organization; 2004.
27. Clinical and Laboratory Standards Institute (CLSI). *Protection of Laboratory Workers from Occupationally Acquired Infections; Approved Guideline—Third Edition*. CLSI Document M29-A3. Wayne, PA: CLSI; 2005.
28. Schrott RW, Foon KA, Beatty SM, et al. Human anti-murine immunoglobulin responses in patients receiving monoclonal antibody therapy. *Cancer Res* 1985;45(2):879-885.
29. Primus FJ, Kelley EA, Hansen HJ, et al. "Sandwich"-type immunoassay of carcinoembryonic antigen in patients receiving murine monoclonal antibodies for diagnosis and therapy. *Clin Chem* 1988;34(2):261-264.
30. Boscato LM, Stuart MC. Heterophilic antibodies: a problem for all immunoassays. *Clin Chem* 1988;34(1):27-33.
31. National Committee for Clinical Laboratory Standards (NCCLS). *Evaluation of Precision Performance of Clinical Chemistry Devices; Approved Guideline*. NCCLS Document EP5-A. Wayne, PA: NCCLS; 1999.
32. National Committee for Clinical Laboratory Standards (NCCLS). *Evaluation of the Linearity of Quantitative Measurement Procedures: A Statistical Approach; Approved Guideline*. NCCLS Document EP6-A. Wayne, PA: NCCLS; 2003.
33. National Committee for Clinical Laboratory Standards (NCCLS). *Interference Testing in Clinical Chemistry; Approved Guideline*. NCCLS Document EP7-A. Wayne, PA: NCCLS; 2002.

ARCHITECT and Chemiflex are trademarks of Abbott Laboratories in various jurisdictions. All other trademarks are property of their respective owners.

 Abbott Laboratories
Diagnostics Division
Abbott Park, IL 60064
USA

 **ABBOTT**
Max-Planck-Ring 2
65205 Wiesbaden
Germany
+49-6122-580



PRODUCED FOR ABBOTT BY



























Fisher Diagnostics
a division of Fisher Scientific Company LLC
a part of Thermo Fisher Scientific Inc.
8365 Valley Pike, Middletown, VA 22645-1905 USA

**Customer Service: Contact your local representative
or find country-specific contact information on
www.abbottdiagnostics.com**

Revised February 2015.
©2005, 2015 Abbott Laboratories



Key to Symbols

	Consult instructions for use
	Manufacturer
	Sufficient for
	Temperature limitation
	Use by/Expiration date
	Assay Diluent
	Conjugate
	Control Number
	Authorized Representative in the European Community
	<i>In Vitro</i> Diagnostic Medical Device
	Lot Number
	Microparticles
	Pre-Trigger Solution
	Produced for Abbott by
	Product of USA
	Reaction Vessels
	Reagent Lot
	List Number
	Replacement Caps
	Sample Cups
	Septum
	Serial number
	Trigger Solution
	Warning: Causes serious eye irritation.
	Warning: Reproductive Hazard
	Wash Buffer